METHOD FOR INHIBITING CYCLOOXYGENASE AND INFLAMMATION USING CHERRY BIOFLAVONOIDS

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U.S. Cl. 514/886; 514/887; 424/195.1

Field of Search 514/886, 887, 514/439, 195.1, 783; 424/195.1

References Cited
4,297,220 10/1981 Meitzner
4,439,458 3/1984 Puri
5,266,685 11/1993 Garbutt
5,665,783 9/1997 Katzavian et al.
5,817,354 10/1998 Mozaffar

Primary Examiner—Jon P. Weber
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ABSTRACT
A method for inhibiting cyclooxygenase enzymes and inflammation in a mammal using a cherry or cherry anthocyanins, bioflavonoids and phenolics is described. In particular a mixture including the anthocyanins, the bioflavonoids and the phenolics is described for this use.

8 Claims, 8 Drawing Sheets.
OTHER PUBLICATIONS


Sigma–Aldrich—Amberlite XAD–2, 500G specifications.


* cited by examiner
Anthocyanin 1
(cyanidin-3-glucosylrutinoside)

Anthocyanin 2
(cyanidin-3-rutinoside)

Anthocyanin 3
(cyanidin-3-glucoside)

FIG. 1
Compound  
Quercetin  
Kaempferol  
Luteolin  
Quercitrin  
Kaempferol 3-rutinoside  
3'-methoxy kaempferol 3-rutinoside  
5,8,4'-trihydroxyl-6,7-dimethoxyflavone

\[
\begin{array}{cccc}
R_2 & R_3 & R_5 & R_6 & R_7 \\
\text{OH} & \text{OH} & \text{H} & \text{OH} & \text{H} \\
\text{H} & \text{OH} & \text{H} & \text{OH} & \text{H} \\
\text{OH} & \text{H} & \text{H} & \text{OH} & \text{H} \\
\text{OH} & \text{rhamnose} & \text{H} & \text{OH} & \text{H} \\
\text{H} & \text{rutinose} & \text{H} & \text{OH} & \text{H} \\
\text{OMe} & \text{rutinose} & \text{H} & \text{OH} & \text{H} \\
\text{H} & \text{H} & \text{OMe} & \text{OMe} & \text{OH} \\
\end{array}
\]

FIG. 2
FIG. 3
FIG. 5

CHERRIES

BLEND WITH WATER
FILTER/CENTRIFUGE

CHERRY SKIN

CHERRY JUICE

XAD-4 OR XAD-16
ELUTE WITH WATER

CHERRY BY-PRODUCT

FOOD/FERMENTATION

XAD-4 OR XAD-16
ELUTE WITH ETHANOL
AND DRY

DRIED CHERRY

CHERRY TABLETS

FIG. 6

RESIN BEADS

10

11

12

13

14

15
**FIG. 7**

- Naproxen
- Cyanidin
- Ibuprofen
- Aspirin

**FIG. 8**

- PGHS-1
- PGHS-2
FIG. 10

FIG. 11
1 METHOD FOR INHIBITING CYCLOOXYGENASE AND INFLAMMATION USING CHERRY BIOFLAVONOIDs

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a continuation-in-part of U.S. Ser. No. 09/317,310, filed May 24, 1999. This application is based upon Provisional application Ser. No. 60/111,945, filed Dec. 11, 1998. This application is also based upon U.S. Provisional Application Ser. No. 60/120,178, filed Feb. 16, 1999.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT
None

BACKGROUND OF THE INVENTION

1. Summary of the Invention

The present invention relates to a method of use of at least one compound isolated from cherries as cyclooxygenase (COX-1 and COX-2) inhibitors. In particular, the present invention provides a natural cherry composition containing a mixture of anthocyanins, bioflavonoids and phenolics for use as anti-inflammatory agents as a result of inhibition of the cyclooxygenase enzymes.

2. Description of Related Art

Many plant-derived compounds may also impart important positive pharmacological or “nutraceutical/phytocidal” traits to foods by way of their abilities to serve as antioxidants by maintaining low levels of reactive oxygen intermediates, as anti-inflammatory agents by inhibiting prostaglandin synthesis, or as inhibitors of enzymes involved in cell proliferation. These activities may be important in ameliorating chronic diseases including cancer, arthritis, and cardiovascular disease (Kinsella et al., Food Tech. 85–89 (1993)). Thus, with natural products, the dietary supplement/food industry and nutraceutical/phytocidal companies have the opportunity to employ compounds which can not only enhance food stability as effectively as synthetic antioxidants, but can also offer significant health benefits to the consumer.

Cherries are thought to have beneficial health properties in general. Consumption of cherries was reported to alleviate arthritic pain and gout (Hame, P. B., et al. Cherokee Plants 28: Herald: Raleigh, N. C. (1975)) although there is no evidence for its active components or mode of action. These beneficial effects may be partially associated with the abundance of anthocyanins, the glycosides of cyanidin.

Prunus Cerasus L. (Rosaceae), cv. MONTMORENCY is the major tart cherry commercially grown in the United States. In order to challenge the MONTMORENCY monoculture, a new cultivar, BALATON tart cherry (Uferberio ferrus), was introduced into the United States in 1964, and has been tested in Michigan, Utah, and Wisconsin. BALATON produces fruits darker than MONTMORENCY.

Colorants like anthocyanins have been regarded as the index of quality in tart cherries. Most importantly, recent results showed that anthocyanins such as cyanidin-3-glucoside have strong antioxidant activities (Buda, T., et al., J. Agric. Food Chem. 42:2407–2410 (1994)). Early studies have showed that MONTMORENCY cherry contains the anthocyanins cyanidin-3-gentiobioside and cyanidin-3-rutinoside (Li, K. C., et al., J. Am. Chem. Soc. 78:979–980 (1956)). Cyanidin-3-glucosylrutinoside was also found in six out of the seven sour cherry varieties (Harborne, J. B., et al., Phytochemistry 3:453–463 (1964)). Dekazos (Dekazos, E. D., J. Food Sci. 35:237–241 (1970)) reported anthocyanin pigments in MONTMORENCY cherry as peonidin-3-rutinoside peonidin and cyanidin along with cyanidin-3-sophoroside, cyanidin-3-rutinoside and cyanidin-3-glucoside. However, cyanidin-3-glucosylrutinoside as well as cyanidin-3-glucoside, cyanidin-3-sophoroside and cyanidin-3-rutinoside were identified as main pigments in sour cherries. Using HPLC retention values, Chandra et al (Chandra, A., et al., J. Agric. Food Chem. 40:967–969 (1992)) reported that cyanidin-3-sophoroside and cyanidin-3-glucoside were the major and minor anthocyanins, respectively, in Michigan grown MONTMORENCY cherry. Similarly, cyanidin-3-xyllosylrutinoside was detected as a minor pigment in MONTMORENCY cherry (Shrikhande, A. J. and F. J. Francis, J. Food Sci. 38:649–651 (1973)).

In the prior art, production of pure anthocyanin (compounds 1–3 of FIG. 1) from BALATON and MONTMORENCY cherry juices was carried out first by adsorbing the pigment on an AMBERLITE XAD-2 (Sigma Chemicals) column (Chandra, A., et al., J. Agric. Food Chem. 41:1062–1065 (1993)). The column was washed with water until the eluant gave a pH of approximately 7.0. The adsorbed pigments along with other phenolics were eluted with MeOH. The resulting crude anthocyanins were fractionated and purified by C-18 MPLC and HPLC, respectively, to afford pure anthocyanins for spectral studies. Purification of 500 mg crude MONTMORENCY anthocyanins from AMBERLITE XAD-2 yielded 60 mg of pure anthocyanins 1–3 compared to 391.43 mg from BALATON. This research indicated that crude anthocyanins from MONTMORENCY obtained from the XAD-2 contained a high percentage of other organic compounds. There was no attempt to use the crude mixture of phenolics and anthocyanins for any purpose. U.S. Pat. Nos., 5,266,685 to Garbutt, 5,665,783 to Katsakian et al and 5,817,354 to Mozaffar describe various adsorbent resins and their use for isolating unrelated products. These patents are only illustrative of the general state of the art in the use of adsorbent resins.

122 2062–2068 (1982); and Baumann, J., et al., Prostaglandins 20 627–640 (1980)). The C₂₇-C₃₀ double bond, which determines the coplanarity of the hetero rings appears to be a major determinant of COX activity (Wurm, G., et al., Deutscher Apotheker Zeitung 122 2062–2068 (1982)). Certain prenylated flavonoids, such as morusin, were also active, because of their higher lipophilicity (Kimura, Y., et al., Chem. Pharm. Bull. 34 1223–1227 (1986)). Also, unsubstituted flavone is a good COX inhibitor (Mower, R. L., et al., Biochem. Pharmacol. 33 357–364 (1984); and Welton, A. F., et al., Prog. Clin. Biol. Res. 213 231–242 (1986)). Most of the flavanones studied in the past did not show significant COX inhibition, except for the flavanone-3-ol, silibinin (Kalkbrenner, F., et al., Pharmacology 44 1–12 (1992)). However, the structure-activity relationships of isoflavonoids are not reported.

There is a need for natural product derived compositions for use as cyclooxygenase inhibitors and as anti-inflammatory agents.

SUMMARY OF THE INVENTION

The present invention relates to a method for inhibiting cyclooxygenase or prostaglandin H synthase enzymes which comprises: providing at least one compound isolated from a cherry with at least one of the enzymes to inhibit the enzymes.

Further, the present invention relates to a method for inhibiting cyclooxygenase or prostaglandin H synthase enzymes which comprises: providing at least one flavonoid compound isolated from a cherry with at least one of the enzymes to inhibit the enzymes.

Further, the present invention relates to a method for inhibiting inflammation in a mammal which comprises: administering at least one compound isolated from a cherry to the mammal to inhibit inflammation.

Further, the present invention relates to a method for inhibiting inflammation in a mammal which comprises: administering at least one flavonoid, anthocyanin or phenolic compound isolated from a cherry to the mammal to inhibit the inflammation.

Finally, the present invention relates to a method for inhibiting inflammation in a mammal which comprises administering cyanidin to the mammal to inhibit inflammation.

The term “anthocyanins” includes the color producing compounds contained in cherries. For the purpose of this application this includes the aglycone cyanidin.

The term “bioflavonoids” means the isoflavonoid and flavonoid compounds contained in cherries.

The term “phenolics” refers to compounds with a phenyl group and having one or more hydroxyl groups.

The compounds isolated from cherries are most useful with living material. The living material can be in an animal or human. It can also be in tissue culture.

OBJECTS

It is therefore an object of the present invention to provide a cherry compound which can be used as cyclooxygenase inhibitors and anti-inflammatory agents. Further, it is an object of the present invention to provide a method for isolating the composition on a commercial scale. Finally, it is an object of the present invention to provide a natural source compound which is economical to prepare and easy to use. These and other objects will become increasingly apparent by reference to the following description and the drawings.

DESCRIPTION OF THE DRAWINGS

FIG. 1 shows the structure of the isolated anthocyanins (colorants) from BALATON and MONTMORENCY cherries. The aglycone cyanidin has a hydroxyl group at position 3.

FIGS. 2 and 3 are drawings showing the major bioflavonoids isolated from the cherries.

FIG. 4 shows the phenolics isolated from tart cherries.

FIG. 5 shows the steps in the method of producing the preferred isolate as described in Examples 1 and 2.

FIG. 6 is a schematic drawing showing the use of an open vessel 10 for holding resin beads, which remove anthocyanins, bioflavonoids and phenolics from the cherry juice.

FIG. 7 is a dose-response curve for the inhibition of the human PGHS-1 enzyme by cyanidin. The antiinflammatory activity of cyanidin was estimated by its ability to inhibit the cyclooxygenase activity of the PGHS-1 enzyme. Cyanidin gave an I₅₀ value of 90 μM for PGHS-1 enzyme, while the NSAID aspirin, naproxen, and ibuprofen gave I₅₀ values of 1050, 11, and 25 μM, respectively.

FIG. 8 is a dose-response curve for the inhibition of PGHS-1 and PGHS-2 enzymes by cyanidin. Cyanidin gave I₅₀ values of 90 and 60 μM for PGHS-1 and PGHS-2 enzymes, respectively.

FIG. 9 is a graph showing the inhibitory effect of PGHS-1 (COX-1) by flavonoids and isoflavonoids at 200 μM concentrations. Data is expressed as mean ±S.E. of triplicate.

Kämpferol 3-rutinoside, 3′-methoxy kaempferol 3-rutinoside, 5,8,4′-triarylhydroxy-6,7-dimethoxyflavone and quercetin were not active at 1000 μM concentrations.

FIG. 10 is a graph showing dose response curves for the inhibition of the PGHS-1 enzyme (COX-1) by flavonoids from BALATON tart cherries compared to the non-steroidal anti-inflammatory drugs, naproxen, aspirin, and ibuprofen. The I₅₀ of kaempferol, quercetin, luteolin, aspirin, naproxen and ibuprofen are 180, 350, 300, 1050, 11 and 25 μM, respectively. Data is expressed as mean ±S.E. of triplicate.

FIG. 11 are graphs showing dose response curve for the inhibition of the PGHS-1 enzyme (COX-1) by isoflavonoids from BALATON tart cherries compared to the non-steroidal anti-inflammatory drugs, naproxen, aspirin and ibuprofen. The I₅₀ of daidzein, biochanin A, genistein, aspirin, naproxen and ibuprofen are 400, 350, 80, 1050, 11 and 25 μM, respectively. Data is expressed as mean ±S.E. of triplicate.

DESCRIPTION OF PREFERRED EMBODIMENTS

The isolates are preferably prepared as a mixture of anthocyanins, bioflavonoids and phenolics by a method for producing a mixture comprising anthocyanins, bioflavonoids and phenolics from cherries as a composition which comprises:

(a) providing an aqueous solution containing the anthocyanins, bioflavonoids and phenolics from the cherries;

(b) removing the anthocyanins, bioflavonoids and phenolics onto a resin surface from the aqueous solution;

(c) eluting the resin surface with an eluant to remove the anthocyanins, bioflavonoids and phenolics from the resin surface; and

(d) separating the eluant from the anthocyanins, bioflavonoids and phenolics.
The cherries used to produce the isolates can be sweet or sour. Tart cherries contain high levels of malic acid in addition to other organic acids which contribute to the sour taste of tart cherries. The method isolates malic acid and other organic acids containing sugars which can be used in foods to provide tartness and flavor. Most preferred are the BALATON and MONTMORENCY cherries.

The isolated mixture of anthocyanins, bioflavonoids and phenolics can be tableted and used as a natural antacutecial, phytocutecial or dietary supplement. In general, the tablets provide a daily dose of the anthocyanins and bioflavonoids of about 1 to 200 mg, preferably a daily dose of 10–100 mg. One hundred (100) cherries provide 10 to 100 mg of anthocyanins and bioflavonoids. The phenolics (FIG. 4) are provided in an amount of 0.1 to 100 mg as a daily dose. One hundred cherries provide 1–50 mg of phenolics. The amount of the anthocyanins, bioflavonoids and phenolics can be adjusted by isolating the individual compounds and blending them together. It is preferred to use the natural mixture of the anthocyanins, bioflavonoids and phenolics which is isolated by the method using the adsorbent resin.

The resin has a surface to which the anthocyanins, bioflavonoids and the phenolics are adsorbed. A preferred class of adsorbents are polymeric crosslinked resins composed of styrene and divinylbenzene such as, for example, the AMBERLITE series of resins, e.g., AMBERLITE XAD-4 and AMBERLITE XAD-16, which are available commercially from Rohm & Haas Co., Philadelphia, Pa. Other polymeric crosslinked styrene and divinylbenzene adsorptive resins suitable for use according to the invention are XE-4257, XE-4022, XUS-40323 and XUS-4032 manufactured by the Dow Chemical Company, Midland, Mich., and the like.

It is preferred to use commercially available, FDA-approved, styrene-divinyl-benzene (SDVB) cross-linked copolymer resin, (e.g., AMBERLITE XAD-16). Thus, in the preferred embodiment, AMBERLITE XAD-16, commercially available from Rohm and Haas Company, and described in U.S. Pat. No. 4,297,220, herein incorporated by reference, is used as the resin. This resin is a non-ionic hydrophobic, cross-linked polystyrene divinyl benzene adsorbent resin. AMBERLITE XAD-16 has a macrotetecular structure, with both a continuous polymer phase and a continuous porous phase. In a particularly preferred embodiment, the resin used in the present invention has a particle size ranging from 100–200 microns.

It is contemplated that other adsorbents such as those in the AMBERLITE XAD adsorbent series which contain hydrophobic macrotetecular resin beads, with particle sizes in the range of 100–200 microns, will also be effective in the methods of the present invention. Moreover, different variations of the AMBERLITES, such as the AMERICHROM CG series of adsorbents, used with particle sizes in the range of 100–200 microns, may also be suitable for use in the present invention. The AMBERLITE XAD-16 is preferred since it can be re-used many times (over 100 times). However, it is contemplated that for food, the use of governmentally approved resins in the present invention will be considered important and/or desirable.

Any solvent can be used to remove the adsorbed anthocyanins, bioflavonoids and phenolics. Preferred are lower alkanols containing 1 to 4 carbon atoms and most preferred is ethanol (ethyl alcohol) since it is approved for food use. Typically the ethanol is azeotroped with water; however, absolute ethanol can be used. Water containing malic acid and sugars in the cherries pass through the column. These are collected and can be used in foods as flavors.

The anthocyanins, bioflavonoids and phenolics are preferably isolated from the BALATON and the MONTMORENCY cherries. The composition of the cherries is in part shown in part by U.S. application Ser. No. 08,799,788 filed Feb. 12, 1997 and in part U.S. application Ser. No. 60/111,945, filed Dec. 11, 1998 and 60/120,178, filed Feb. 16, 1999, which are incorporated by reference herein.

The term “carrier” or “bulking agent” is used to mean a composition which is added to increase the volume of the composition of the purified composition from the cherry. Preferred is dried cherry pulp. These include any edible starch containing material, such as the compounds from BALATON and MONTMORENCY cherries. Juiced cherry tissue was sequentially extracted with hexane, ethyl acetate and methanol. Both methanol and ethyl acetate fractions showed strong antioxidant activity in the screening assay. The ethyl acetate fraction was further purified by silica gel vacuum liquid chromatography to yield four subfractions; the subfraction was further separated into seven fractions by preparative reverse phase HPLC. Figs. 2 and 3 show the bioflavonoids isolated from the BALATON cherries. There are thus numerous analogous or homologous compounds in the tart cherries. The anthocyanin components obtained from the juice fraction also have been identified and fully characterized (Chandra, A., et al., J. Agric. Food Chem. 41:1062 (1992); Wang, H., et al., J. Agric. Food Chem. 45:2556–2560 (1997)) and for rapid screening of antioxidant activity (Arora, A. and G. M. Strasburg, J. Agric. Food Chem. 44:1031–1040 (1996)). These methods are being utilized to identify the characteristics from BALATON and MONTMORENCY cherries. Juice cherry tissue was sequentially extracted with hexane, ethyl acetate and methanol. Both methanol and ethyl acetate fractions showed strong antioxidant activity in the screening assay. The ethyl acetate fraction was further purified by silica gel vacuum liquid chromatography to yield four subfractions; the subfraction was further separated into seven fractions by preparative reverse phase HPLC. Figs. 2 and 3 show the bioflavonoids isolated from the BALATON cherries. There are thus numerous analogous or homologous compounds in the tart cherries. The anthocyanin components obtained from the juice fraction also have been identified and fully characterized (Chandra, A., et al., J. Agric. Food Chem. 41:1062 (1992); Wang, H., et al., J. Agric. Food Chem. 45:2556–2560 (1997)).

Two novel phenolic compounds were identified: I) 1-(3'-4'-dihydroxy cinnamoyl)-2,3-dihydroxy cyclopetanone, and II) 1-(3'-4'-dihydroxy cinnamoyl)-2,5-dihydroxy cyclopetanone. Other compounds isolated from the ethyl acetate extract of cherry fruits and characterized by spectral methods include: 1-(3'-methyl, 4'-hydroxy cinnamoyl) quinic acid, 2-hydroxy-3-(2'-hydroxyphenyl) propanoic acid, methyl 2-hydroxy-3-(2'-hydroxyphenyl) propanoate, D(+)-malic acid, β-sitosterol and β-β-sitosterol glucoside. FIG. 4 shows some of the phenolics which were isolated.

EXAMPLES 1 and 2

As shown in FIG. 5, individual quick frozen (IQF) cherries (which had been pitted) were defrosted and blended in an industrial WARING blender. The mixture was centrifuged at 10,000 rpm and the juice was decanted. The residue, pulp, was further pressed with cheese cloth to remove any additional juice.
The pulp was lyophilized at 15°C. The juice was processed on AMBERLITE XAD-16 HP resin to produce cherry sour, anthocyanins, bioflavonoids and phenolics. The XAD-16 resin, 1 kg, was washed with ethanol (1-2 L) and then washed with water (6 L). The XAD-16 resin was allowed to stand in water for 1 hour before loading into a glass column (10 IDx90 cm long) with a cotton plug. The packed column was washed with water (2 L) before loading the juice for separation. 800 mL juice was purified each time. The juice was added onto the surface of the column and allowed to settle with no flow. It was then eluted with water and the first 1 L was discarded. The next 2 L of washing was collected, since it contained the cherry juice which was sour since it contained malic acid and sugars from the cherries. The column was then washed with an additional 4 L of water in the case of BALATON and 5 L for MONTMORENCY cherry juice. Once the cherry juice was collected, the remainder of the washing with water were discarded. The column was then eluted with ethanol (1.3-1.5 L) and collected the red solution containing anthocyanins, bioflavonoids and phenolics (700-800 mL). The column was then run dry and washed with 10 L of water before repeating the process many of times (over 100).

The red alcoholic solution was then evaporated under vacuum a (20 millitorr) to remove ethanol and the aqueous solution, stabilized with 50 ppm ascorbic acid, was lyophilized at 10°C. The red powder was collected and stored.

Example 1 results:

<p>| | |</p>
<table>
<thead>
<tr>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>BALATON cherry</td>
<td></td>
</tr>
<tr>
<td>Weight of IQF cherries</td>
<td>15.74 kg</td>
</tr>
<tr>
<td>Weight of dried pulp</td>
<td>605 g</td>
</tr>
<tr>
<td>Volume of juice</td>
<td>12.16 L</td>
</tr>
<tr>
<td>Weight of anthocyanins, bioflavonoids and phenolics (red powder)</td>
<td>31.35 g</td>
</tr>
<tr>
<td>Volume of sour byproduct (malic acid and sugars) @ 35 L</td>
<td></td>
</tr>
</tbody>
</table>

Example 2 results:

<p>| | |</p>
<table>
<thead>
<tr>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>MONTMORENCY cherry</td>
<td></td>
</tr>
<tr>
<td>Weight of IQF cherries</td>
<td>30.45 kg</td>
</tr>
<tr>
<td>Weight of dried pulp</td>
<td>895 g</td>
</tr>
<tr>
<td>Volume of juice</td>
<td>24.03 L</td>
</tr>
<tr>
<td>Weight of anthocyanins, bioflavonoids and phenolics (red powder)</td>
<td>47 g</td>
</tr>
<tr>
<td>Volume of cherry by-product (malic acid and sugars) @ 75 L</td>
<td></td>
</tr>
</tbody>
</table>

The red powders of Examples 1 and 2 were preferably mixed with dried pulp as a carrier and tabletted into 1 to 1000 mg tablets including the carrier (1 adult daily dose).

Various food grade acids can be added to the isolated anthocyanins, bioflavonoids and phenolics to prevent decomposition. Preferably they do not add flavor. Ascorbic acid (vitamin C) is preferred. The acid can be added before or after, preferably before drying of the cherry compounds.

For small scale processing, lyophilization is used to remove water. For larger scale production, drying in an air circulating oven is preferred.

EXAMPLE 3

As shown in FIG. 6, an open vessel 10 is provided with an inlet line 11 and an outlet line 12, with valves 13 and 14, respectively. The resin beads 15 are provided in the open vessel 10. Water is introduced into the vessel 10 and then removed through outlet line 12 and discarded. The cherry juice (without the pulp or pits) as in Example 1 is introduced to the vessel 10 and allowed to stand for 25 minutes. The temperature of the water and juice is between about 20 and 30°C. The cherry juice residue containing malic acid and sugars is then removed through the outlet line 12 and retained as a food flavoring. The resin 15 in the vessel is then washed again with water from inlet line 11 and then removed and discarded through outlet line 12. The anthocyanins, bioflavonoids and phenolics on the resin particles are then extracted using 95% ethanol introduced through inlet line 11. The ethanol containing the anthocyanins, bioflavonoids and phenolics is removed from the vessel 10. The ethanol is removed from the anthocyanins, bioflavonoids and phenolics and dried using flash drying under nitrogen. The resulting powder is preferably then mixed with dried cherry pulp or other carrier as in Example 1. The resin particles are washed with water and then the resins and ethanol are recycled many times.

EXAMPLE 4

The antiinflammatory assays on the anthocyanins and cyanidin were conducted using prostaglandin endoperoxide H synthase-1 and -2 isozymes (PGHS-1, and -2) and were based on their ability to convert arachidonic acid to prostaglandins (PGs). The positive controls used in this experiment were aspirin, naproxen, and ibuprofen. Aspirin gave an IC50 value of 1050 μM each against PGHS-1 and PGHS-2 enzymes (FIG. 7). Naproxen and ibuprofen gave IC50 values of 11 and 25 nM against PGHS-1 enzyme, respectively (FIG. 7). A preliminary experiment with the mixture containing anthocyanins 1–3 (FIG. 1) showed PGHS-1 and PGHS-2 activities at 33 ppm concentration. The aglycon cyanidin showed good PGHS-1 and -2 inhibitory activities, with IC50 values of 90 and 60 nM, respectively (FIGS. 7 and 8). The ratio of IC50 values for PGHS-1 to PGHS-2 was about 0.56 (FIG. 8). However, pure anthocyanins 1–3 showed little or no activity against PGHS-1 and PGHS-2 at 300-nM test concentrations. Higher concentrations of anthocyanins 1 and 2, on the contrary, increased the activity of enzyme. This is probably due to the ability of anthocyanins 1 and 2 to act as oxygen carriers at high concentration and enhance the oxygen uptake. It is noted that anthocyanins are hydrolyzed in the gut of a mammal to cyanidin and other compounds and thus effective in vivo.

For measurements of time-dependent inhibition of PGHS-2 enzyme activity by cyanidin, the enzyme was preincubated at 37°C with 15 nM of cyanidin (one-fourth of the concentration of IC50) and added to an oxygen electrode chamber with arachidonic acid substrate to initiate the reaction. The results suggest that the rate of inhibition of PGHS-2 did not change with time.

The specific inhibition of the PGHS-2 enzyme is a major advance in antiinflammatory therapy because it significantly reduces the adverse effects of nonsteroidal antiinflammatory drugs (NSAIDs). It is generally believed that ulcerogenic and other adverse properties of NSAIDs result from the inhibition of PGHS-1, whereas the therapeutically desirable effects come from the inhibition of PGHS-2 enzyme.

Similarly, cyanidin showed better antiinflammatory activity than aspirin in the inflammatory assays. The antioxidant and antiinflammatory properties of anthocyanins and cyanidin suggest that consumption of cherries may have the potential to reduce cardiovascular or chronic diseases in humans.
In particular, arachidonic acid and a microsomal fraction of ram seminal vesicles containing PGHS-1 enzyme suspended in 100 mM Tris pH 7.8 and 300 μM diethylthiobis-carbamic acid (DDC) as a preservation were purchased from Oxford Biomedical Research (Oxford, Mich.). Recombinant human PGHS-2 enzyme was initially obtained from Dr. David Dewitt (Department of Biochemistry, Michigan State University, East Lansing, Mich.) and then purchased from Oxford Biomedical Research (Oxford, Mich.). Naproxen, ibuprofen, and hemoglobin were purchased from Sigma Chemical Co. (St. Louis, Mo.). Anthocyanins 1–3 were purified from Balaton tart cherry by HPLC and were identified by 1H and 13C NMR spectral data.

To prepare cyanidin, the anthocyanin mixture containing 1–3 (FIG. I, 500 mg) was stirred with 3N HCl (20 mL) at 80°C for 10 hours. The reaction mixture was purified on a XAD-4 column as in the preparation of anthocyanins. The MeOH solution of cyanidin was evaporated to dryness to yield a red amorphous powder (190 mg) and stored at -30°C until use.

In the antiinflammatory assay, cyclooxygenase activities were measured by using PGHS-1 enzyme (ca. 5 mg protein/ml in 0.1 M TrisHCl, pH 7.8), a homogeneous protein purified from ram seminal vesicles. Microsomal preparations from recombinant human prostaglandin synthase-2 (COX-2) were obtained from insect cell lysate. Assays were performed at 37°C by monitoring the initial rate of O2 uptake using an O2 electrode (Yellow Springs Instrument Inc., Yellow Springs, Ohio). Each assay mixture contained 3 mL of 0.1 M TrisHCl, pH adjusted to 7 by the addition of 6M HCl, 1 mM phenol, 85 μg hemoglobin, and 10 μM of arachidonic acid. Reactions were initiated by the addition of 5–25 μg of microsomal protein in a volume of 15–50 μL.

Instantaneous inhibition of enzyme activity was determined by measuring the cyclooxygenase activity initiated by adding aliquots of microsomal suspensions of PGHS-1 or PGHS-2 (10 μM O2/min cyclooxygenase activity/ aliquot) to assay mixtures containing 10 μM arachidonate and various concentrations of the test substances (10–1100 μM). The IC50 values represent the concentrations of the test compound that gave half-maximal activity under the standard assay conditions.

**EXAMPLE 5**

This is an antiinflammatory assay for cyclooxygenase inhibition activity of flavonoids and isoflavonoids. Arachidonic acid and microsomal suspensions of PGHS-1 (COX-1) and COX-2 (PGHS-2) were purchased from Oxford Biomedical Research (Oxford, Mich., USA). Genistein, genistin, naringenin, quercetin, 5,6,4-trihydroxy-6,7-dimethoxyflavone, kaempferol-3-rutinoside and 3-methoxy kaempferol 3-rutinoside were purified from BALATON tart cherry by HPLC and were identified by 1H- and 13C NMR spectral data. Daidzein and formononetin were purchased from Research Plus, Inc. (Bayonne, N.J., USA). Biochanin A, kaempferol, quercitin, naproxen, ibuprofen and hemoglobin were purchased from Sigma Chemical Co. (St. Louis, Mo., USA). Luteolin was purchased from Adams Chemical Co. (Round Lake, Ill., USA).

For measuring the COX activity, flavonoids or isoflavonoids were dissolved in DMSO to yield 40 mM stock solution and was further diluted in the desired concentration according to the COX-1/COX-2 inhibitory activity of each compound assayed.

Antioxidative assay: COX activities were measured using microsomal suspensions of PGHS-1 and PGHS-2.
activity of kaempferol and quercetin were reported in other model systems (Kalkbrenner, F., et al., Pharmacology 44 1–12 (1992); Hoult, J. R. S., et al., Agents and Actions 42 787–792 (1988); and Moroney, M. A., et al., J. Pharm. Pharmacol. 40 787–792 (1988)). The OH group at C_3 position is also important for the activity. However, the glycosylation of the OH group at C_3 decreased the activity considerably. Comparing the COX-1 inhibitory activity of flavones (luteolin) with their corresponding flavanoids (quercetin), it can be concluded that the absence of an OH group at C_3 enhanced the COX-1 activity slightly. It is important to note that quercetin 3-rhamnoside was not active in the assay, but reported to have in vivo anti-inflammatory activity (Sanchez de Medina, L. H., et al., J. Pharmacol. Exp. Ther. 278 771–779 (1996)). This may be due to the in vivo metabolism of quercetin 3-rhamnoside to quercetin. The C_2–C_3 double bond, which determines the coplanarity of the hetero-rings in flavonoids and isoflavonoids, was essential for a higher COX inhibitory activity. If the double bond was saturated, the COX-1 inhibitory effect was dramatically decreased as in the case of naringenin (Fig. 9). This result is consistent with previous reports (Wurm, G., et al., Deutche Apotheker Zeitung 122 2062–2068 (1982); Kalkbrenner, F., et al., Pharmacology 44 1–12 (1992)). Also, the multiple substituents such as OH and OMe groups in the A ring of the flavonoids caused little or no COX-1 inhibition as demonstrated by the activity of 5,8,4′-tri hydroxy-6,7-dimethoxy flavone.

Among the isoflavonoids (Figs. 2 and 3), genistein showed the highest COX-1/COX-2 inhibitory activity. The activity was dramatically decreased in genistein, when the 7-OH group in ring A of genistein was glycosylated. Also, the hydroxyl group at C-4′ in isoflavonoids is essential for the COX-1/COX-2 inhibitory activity. When 4′-OH groups in genistein and daidzein were methoxylated, the activity decreased considerably. The 5-OH group in isoflavonoids is also important for COX-1/COX-2 inhibitory effect. These results indicated that C_3′, C_4, and C_5 hydroxyl groups in isoflavonoids are essential for COX-1 inhibition. Comparison of genistein with that of kaempferol indicates that substitutions on ring B and at C_3 of ring C enhances COX-1/COX-2 inhibitory effect. In addition to COX-1/COX-2 inhibition, these isoflavonoids and flavonoids also showed good antioxidant activity. Both COX-1 inhibitory and antioxidant activities of these compounds suggests that tart cherries may possess significant health benefits to humans. These bioflavonoids may be partially responsible for the anecdotal claims associated with tart cherries of alleviating pain related to treatment of arthritis and gout.

Thus several flavonoids and isoflavonoids isolated from BALATON tart cherry were assayed for prostaglandin H endoperoxide synthase (PGHS-1 or PGHS-2) enzyme activity. Genistein showed the highest COX-1 inhibitory activity among the isoflavonoids studied with an IC_{50} value of 80 μM. Kaempferol gave the highest COX-1 inhibitory activity among the flavonoids tested with an IC_{50} value of 180 μM. The structure-activity relationships of flavonoids and isoflavonoids revealed that hydroxyl groups at C_3′, C_4, and C_5 in isoflavonoids were essential for appreciable COX-1 inhibitory activity. Also, the C_2–C_3 double bond in flavonoids is important for COX-1 inhibitory activity. However, hydroxyl group at C_3′ position decreased the COX-1/COX-2 inhibitory activity by flavonoids.

**EXAMPLE 6**

The composition of Examples 1 and 2 were tested for anti-inflammatory activity using cyclooxygenase 1 and 2 (COX-1 and COX-2) in an assay as described in Wang et al., J. Nat. Products 62:294–296 (1999); Wang et al., J. of Ag. and Food Chemistry, 47: 840–844 (1999) and Wang et al., J. of Nat. Products, 62:86–88 (1999) and Examples 4 and 5. The results were that the compositions exhibited anti-inflammatory activities, specifically strong inhibition of COX-1 and COX-2.

It is intended that the foregoing description be only illustrative of the present invention and that the present invention be limited only by the hereinafter appended claims.

We claim:

1. A method for inhibiting cyclooxygenase or prostaglandin H synthase enzymes comprising:
   - providing at least one compound anthocyanin selected from the group consisting of cyanidin-3-glucosylrutinoside, cyanidin-3-rutinoside and or cyanidin-3-glucose isolated from the fruit of a cherry to inhibit the enzymes.

2. The method of claim 1 wherein the method is in vitro.

3. The method of claim 1 wherein the method is in vivo.

4. The method of any one of claims 1, 2 or 3 wherein the compound is from a tart cherry.

5. The method of any one of claims 1, 2 or 3 wherein the compound is isolated from a sweet cherry.

6. A method for inhibiting inflammation in a mammal comprising:
   - administering to a mammal in need of such treatment, an effective amount of at least one anthocyanin obtained from the fruit of a cherry, said anthocyanin selected from the group consisting of cyanidin-3-glucosylrutinoside, cyanidin-3-rutinoside, and cyanidin-3-glucose and mixtures thereof to inhibit said inflammation.

7. The method of claim 6 wherein said anthocyanin is obtained from Prunus avium, Prunus cerasus, and mixtures thereof.

8. The method of claim 6 wherein said anthocyanin is obtained from Prunus cerasus.
It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

**Column 3.**
Line 48, “incudes” should be -- includes --.

**Column 7.**
Line 23, “many of times” should be -- many times --.

**Column 9.**
Line 12, “Balaton” should be -- BALATON --.

**Column 11.**
Line 27, “5, 8, 4-trihydroxy” should be -- 5, 8, 4’- trihydroxy --.

**Column 12.**
Line 28, (Claim 1), “compound” after “one” and before “anthocyanin” should be deleted.

Signed and Sealed this
Sixteenth Day of October, 2001

*Attest:*

Nicholas P. Godici

NICHOLAS P. GODICI
Attesting Officer Acting Director of the United States Patent and Trademark Office